66271-02-1; Boc-L-Ser(Bzl)-OCH₂PhCH₂COOH CHA salt, 66271-04-3; Boc-L-Met-OCH₂PhCH₂COOH CHA salt, 66271-06-5; Boc- $L-Lys(Z)-OCH_2C_6H_4-p-CH_2COOH_2COPh, 66271-07-6; Boc-L-As$ p(OBzl)-OCH₂C₆H₄-p-CH₂COOCH₂COPh, 66271-08-7; Boc-L-Ser(Bzl)-OCH₂C₆H₄-p-CH₂COOCH₂COPh, 66271-09-8; Boc-L-Met-OCH₂C₆H₄-p-CH₂COOCH₂COPh, 66271-10-1; Boc-Val-OCH₂C₆H₄-p-CH₂CONHCH₂Ph, 66271-11-2; Boc-L-Val DCHA salt, 16944-17-5; Boc-L-Val-4-(OCH2)PhCH2CO-4-(OCH2)PhCH2COOH, 66271-12-3; N-hydroxysuccinimide, 6066-82-6; 4-(bromomethyl)phenylacetic, acid N-hydroxysuccinimide ester, 66271-13-4; Boc-Valyl-4-(oxymethyl)phenylacetic acid N-hydroxysuccinimide ester, 66271-14-5; 4-(acetoxymethyl)phenylacetic acid, 61165-81-9; Boc-Gly Cs salt, 42538-64-7; Boc-L-Phe Cs salt, 42538-61-4; Boc-L-Val Cs salt, 42538-62-5; Boc-Leu, 13139-15-6; L-Leu-L-Val, 13588-95-9; Boc-Gly, 4530-20-5; Boc-L-Ala, 15761-38-3; Leu-Ala-Gly-Val, 17195-26-5; benzylamine, 100-46-9; p-nitrophenyltrifluoroacetate, 658-78-6; butylamine, 109-73-9; Boc-Valine butylamide, 66271-15-6; Boc-Gly-NHBzl, 19811-52-0.

References and Notes

- (1) Supported in part by Grant AM 01260 from the U.S. Public Health Service and by a grant from the Hoffmann-La Roche Foundation. M. E. was the recipient of a fellowship from the Deutsche Forschungsgemeinschaft.
- (2) Abbreviations used: Boc, tert-butyloxycarbonyl; CHA, cyclohexylamine; DCC, dicyclohexylcarbodiimide; DCHA, dicyclohexylamine; DMF, N,N-dimethylformamide; KelF-g-styrene, radiation-induced graft polymer of styrene on solid poly(trifluorochloroethylene); NMR, nuclear magnetic resonance; Pam, phenylacetamidomethyl; PLC, preparative layer chromatography; R, resin; TLC, thin-layer chromatography. Other nomenclature and symbols follow the Tentative Rules of the IUPAC-IUB Commission on Biochemical Nomenclature, J. Biol. Chem., 241, 2491 (1966); 242, 555
- (1967); 247, 977 (1972).
 (3) R. B. Merrifield, *J. Am. Chem. Soc.*, 85, 2149 (1963).
 (4) B. W. Erickson and R. B. Merrifield, "The Proteins", Vol. 2, 3rd ed, H. Neurath and R. L. Hill, Ed., Academic Press, New York, N.Y., 1976, pp per 5 255-527.
- S. Karlsson, G. Lindeberg, J. Porath, and U. Ragnarsson, Acta Chem. Scand., 24, 1010 (1970). (5)
- (6) U. Ragnarsson, S. Karlsson, and G. Lindeberg, Acta Chem. Scand., 24, 2821 (1970).
- B. Gutte and R. B. Merrifield, *J. Biol. Chem.*, **246**, 1922 (1971).
 P. Fankhauser, B. Schilling, P. Fries, and M. Brenner, in "Peptides-–1971", H. Nesvadba, Ed., North-Holland, Amsterdam, 1973, p 153.
- A. R. Mitchell, B. W. Erickson, M. N. Ryabtsev, R. S. Hodges, and R. B. Merrifield, *J. Am. Chem. Soc.*, **98**, 7357 (1976).
 N. M. Weinshenker and C. M. Shen, *Tetrahedron Lett.*, 3281 (1972).
- H. Ito, N. Takamatsu, and I. Ichikizaki, *Chem. Lett.*, 577 (1975).
 J. T. Sparrow, *J. Org. Chem.*, **41**, 1350 (1976).
 H. E. Zaugg and W. B. Martin, *Org. React.*, **14**, 52 (1965).

- (14) A. R. Mitchell, S. B. H. Kent, B. W. Erickson, and R. B. Merrifield, Tetrahedron Lett., 3795 (1976).
- (15) Occupational Safety and Health Administration, U.S. Department of Labor, *Fed. Regist.*, **39**, 3756 (1974).
- (16) J. C. Sheehan and G. D. Daves, Jr., J. Org. Chem., 29, 2006 (1964) (17) J. Taylor-Papadimitriou, C. Yovanidis, A. Paganou, and L. Zervas, J. Chem. Soc. C, 1830 (1967).
- J. B. Hendrickson and C. Kandall, *Tetrahedron Lett.*, 343 (1970).
 L. Chauffe, L. J. Andrews, and R. M. Keefer, *J. Org. Chem.*, **31**, 3758 (1966).
- (20) M. N. Bogdanov, J. Gen. Chem. USSR (Engl. Transl.), 28, 1670 (1958). In this procedure both chloromethyl methyl ether and bis(chloromethyl) ether are generated. These are potent human carcinogens (see ref 15). (21) K. Suzuki, N. Endo, K. Nitta, and Y. Sasaki in "Proceedings of the 14th
- Symposium on Peptide Chemistry (Japan)", Protein Research Foundation, Osaka, 1977, p 45. B. F. Gisin, *Helv. Chim. Acta*, **56**, 1476 (1973).
- (22)
- (23)M. Bodanszky and S. Natarajan, J. Org. Chem., 40, 2495 (1975).
- (24) Unpublished results of this laboratory.
- (25) Silanizing procedure: B. F. Gisin, personal communication
- (26) J. M. Manning and S. Moore, J. Biol. Chem., 243, 5591 (1968).
 (27) R. S. Hodges and R. B. Merrifield, J. Biol. Chem., 250, 1231 (1975).
 (28) R. B. Merrifield, A. R. Mitchell, and J. E. Clarke, J. Org. Chem., 39, 660
- (1974). (29) O. Schou, D. Bucher, and E. Nebelin, Z. Physiol. Chem., 357, 103
- (1976).
- (30) R. A. Laursen, "Solid-Phase Methods in Protein Sequence Analysis", Pierce
- Chemical Co., Rockford, III., 1975, pp 1–286. (31) J. M. Stewart, *J. Macromol. Sci., Chem.*, **10**, 259 (1976). (32) G. W. Tregear in "Chemistry and Biology of Peptides", J. Meienhofer, Ed.,

- (32) G. W. Iregear in Chemistry and Biology of Peptides, J. Melenhorer, Ed., Ann Arbor Science Publishers, Ann Arbor, Mich., 1972, p 175.
 (33) E. Bayer and M. Mutter, *Nature (London)*, 237, 512 (1972).
 (34) M. Mutter and E. Bayer, *Angew. Chem., Int. Ed. Engl.*, 13, 88 (1974).
 (35) G. Jung, G. Bovermann, W. Göhring, and G. Heusei, In "Peptides— Chemistry, Structure, Biology", R. Walter and J. Meienhofer, Eds., Ann Arbor Science Publishers, Ann Arbor, Mich., 1975, p 433.
 (36) E. Atherton, D. L. J. Clive, and R. C. Sheppard, *J. Am. Chem. Soc.*, 97, 6584
- (1975).
 (37) G. W. Clark, J. Chromatogr., 34, 262 (1968).
 (38) B. F. Gisin and R. B. Merrifield, J. Am. Chem. Soc., 94, 6165 (1972).

- (39) J. A. Patterson in "Biochemical Aspects of Reactions on Solid Supports", G. R. Stark, ed., Academic Press, New York, N.Y., 1971, p 189.
 (40) B. F. Gisin, Anal. Chim. Acta, 58, 248 (1972). (41) J. Scotchler, R. Lozier, and A. B. Robinson, J. Org. Chem., 35, 3151
- 1970). (42) G. W. Anderson, J. E. Zimmerman, and F. M. Callahan, J. Am. Chem. Soc.,
- 86, 1839 (1964).
 (43) H. Yajima, N. Fujii, H. Ogawa, and H. Kawatani, J. Chem. Soc., Chem.

- (45) H. Fajina, N. Fujin, H. Ogawa, and H. Kawadani, J. Orlem. Soc., Chem. Soc., Chem. Commun., 106 (1974).
 (44) S. Sakakibara and N. Inukai, Bull. Chem. Soc. Jpn., 38, 1979 (1965).
 (45) E. Schroder and K. Lübke, Justus Liebigs Ann. Chem., 655, 211 (1962).
 (46) N-(Hydroxymethyl)phthalimide as usually prepared¹³ or commercially supplied has a melting point in the range 137–141 °C and is about 90 mol of the the the the the term for the the the the the term. % pure. This material is satisfactory for use in the procedure described. It can be purified (mp 149.5 °C) via the pyridine complex: E. J. Sakellarios, J. Am. Chem. Soc., 70, 2822 (1948).

Synthesis of Oxysanguinarine

M. Shamma* and H. H. Tomlinson

Department of Chemistry, The Pennsylvania State University, University Park, Pennsylvania 16802

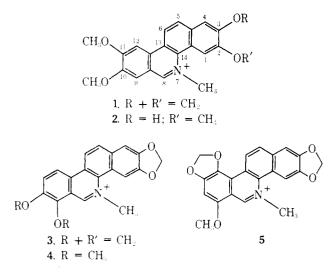
Received January 18, 1978

Base-catalyzed condensation of the homophthalate ester 14 with the imine 15 supplied the lactam amide 16. This compound was saponified to the acid 17 which was homologated by an Arndt-Eistert sequence to the ester 19. Hydrolysis and acid-catalyzed cyclization provided the keto lactam 21. Acid dehydration of the lactam alcohol 22, derived from reduction of 21, was accompanied by air oxidation to provide the desired alkaloid oxysanguinarine (23).

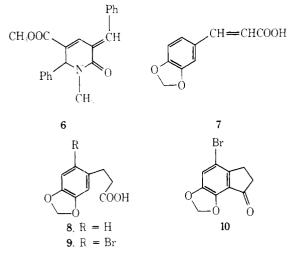
A number of aromatic benzophenanthridine alkaloids possess interesting biological activity. Nitidine (1) and fagaronine (2) have shown anticancer activity,¹ while sanguinarine (3), chelerythrine (4), and chelirubine (bocconine) (5)are nematocides.²

The aim of the present study was to synthesize a naturally occurring aromatic benzophenanthridine, namely oxysanguinarine (23),³ through a route based on the previously reported finding that base-catalyzed condensation of diethyl glutaconate with N-benzylidenemethylamine yields lactam 6.4 The first hurdle was to prepare the homophthalic ester 14, which was to be condensed with piperonylidenemethylamine (15) to afford such lactams as 16, 17, or 18. Homologation of the acid 17 to the acid 20, followed by intramolecular Friedel–Crafts acylation, would then afford keto lactam 21, which would be readily convertible into oxysanguinarine (23).

An eight-step sequence to the homophthalic ester 14 was developed which parallels to some extent, but is superior to, that recorded by Haworth and co-workers for the construction of the corresponding homophthalic acid 13.5 Doebner con-



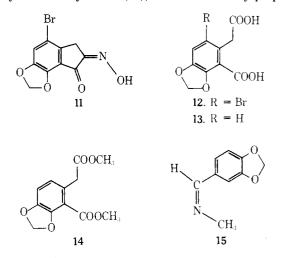
densation of piperonal with malonic acid in refluxing pyridine gave the cinnamic acid 7 in 90% yield. Catalytic hydrogenation of the sodium salt of 7 in water using 5% palladium on carbon furnished, upon acidification, piperonylacetic acid (8) in 95% yield. Attempted cyclodehydration of the bromo acid 9, derived from bromination of 8 in acetic acid, by the Haworth procedure (P_2O_5 in benzene)⁵ involved a tedious workup and resulted in a yield of <20% of the hydrindone 10, so that a



method for achieving this transformation in higher yield was sought. Phosphorus oxychloride, polyphosphoric acid, polyphosphate ester,⁶ super polyphosphoric acid,⁷ and phosphorus oxychloride/zinc chloride⁸ were all tried as cyclizing agents and found to be unsatisfactory. However, when the reaction was run using phosphorus pentoxide in refluxing chlorobenzene under conditions of relatively high dilution, the desired hydrindone **10** was obtained in 40% yield. This material was then nitrosated⁹ using isoamylnitrite to afford the α -oximino ketone **11** in 85% yield.

The second-order Beckmann rearrangement of the α -oximino ketone 11 to the bromo diacid 12 had been reported to proceed in 75% yield.⁵ However, repeated attempts to duplicate this procedure invariably gave yields on the order of 20%, while the reaction workup was troublesome due to formation of emulsions. Several alternative procedures were investigated, and the best method found involved reaction of 11 in a Schotten-Baumann procedure using *p*-toluenesulfonyl chloride and aqueous sodium hydroxide.¹⁰ The transitory nitrile was immediately hydrolyzed by refluxing the strongly basic reaction mixture until the evolution of ammonia had subsided. Upon acidification, the desired bromo diacid 12 was isolated (51%). Debromination with sodium amalgam then afforded the known 3,4-methylenedioxyhomophthalic acid

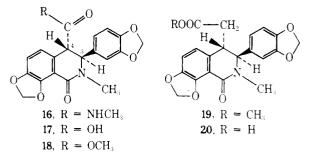
 13^5 in 83% yield, or in 8.3% overall yield from piperonal. Fischer esterification of this diacid gave rise to the corresponding diester 14. The second precursor required for the condensation-cyclization step to the fused lactam was piperonylidenemethylamine (15), which was readily prepared



through condensation of piperonal with methylamine.

Condensation of the diester 14 with the Schiff base 15 required refluxing for a week in a sodium methoxide-methanol solution. A requisite added ingredient for this condensation was methylamine gas, which was passed periodically through the mixture. Lactam 16 was thus isolated in 61% yield. The ¹H NMR spectrum of 16 shows the expected signals for two *N*-methyl groups, two methylenedioxys, and five aromatic protons. Present also are two broad peaks at δ 4.17 and 5.42 for the methine protons at C-4 and C-3, respectively. Both of these peaks are resolved into doublets, $J_{3,4} = 1$ Hz.

Hydrolysis of 16 in 10% aqueous potassium hydroxide afforded the lactam acid 17 in 70% yield. The ¹H NMR spectrum shows only one *N*-methyl signal at δ 3.47, while $J_{3,4} = 0$ Hz, indicating that the dihedral angle between the hydrogens at C-3 and C-4 must be close to 90°.¹¹ The corresponding methyl ester 18 exhibits $J_{3,4} = 1.5$ Hz. Attempts to epimerize the C-4



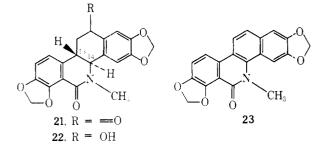
center of 18 with sodium methoxide in methanol gave, as expected, material indistinguishable from 18, so that the molecule must exist in the thermodynamically more stable trans configuration.

Arndt-Eistert homologation of the lactam acid 17 provided ester 19 (60%), which was saponified to the acid 20. The C-3 proton in the ¹H NMR spectrum of 20 appears as a broad singlet with no discernible splitting, which corresponds to a 90° angle between the C-3 and C-4 protons, so that these hydrogens also must be trans to each other.¹¹

A variety of methods for the cyclodehydration of the acid **20** to the ketone **21** were explored, including phosphorus pentoxide in benzene and in chlorobenzene, polyphosphoric acid by itself and in benzene, super polyphosphoric acid,⁷ phosphorus oxychloride, phosphorus oxychloride/zinc chloride,⁸ and polyphosphoric ester in chloroform.⁶ None of these methods proved satisfactory. The cyclodehydration was then

attempted using a mixture of methanesulfonic acid and phosphorus pentoxide.¹² Under these relatively mild conditions, the desired ketone 21 was generated in 44% yield. The ¹H NMR spectrum of this product shows only four aromatic protons, and the signal for H-14 appears as a doublet, $J_{13,14}$ = 11.5 Hz. This large J value is clearly indicative of a trans diaxial hydrogen relationship.¹¹

Sodium borohydride reduction of ketone 21 led to alcohol 22. Dehydration of this compound using *p*-toluenesulfonic



acid was accompanied by air oxidation so that oxysanguinarine (23) was obtained directly from this step. This material was identical with a sample of oxysanguinarine derived from ferricyanide oxidation of sanguinarine (3).¹⁴

The present synthetic method can be readily adapted to the preparation of such alkaloids as nitidine (1) and fagaronine (2), since aromatic benzophenanthridine lactams (oxybenzophenanthridenes) are known to be convertible into aromatic benzophenanthridine salts by reduction with lithium aluminum hydride followed by mercuric acetate oxidation.¹³

Experimental Section

Standard Procedures. All melting points were taken on a melting point block and are uncorrected. Microanalyses were performed by Midwest Microlab, Inc., Indianapolis, Indiana. Thin-layer chromatography was on Brinkmann silica gel F-254 plates (0.25-mm thick). Visualization was accomplished by shining UV light on the plates, or by spraying with chromotropic acid reagent. ¹H NMR spectra are in CDCl₃ with Me₄Si as internal standard unless specified otherwise.

4-Bromo-6,7-methylenedioxy-1-hydrindone (10). A suspension of 73 g (0.51 mol) of P_2O_5 in 2.1 L of chlorobenzene was refluxed with rapid mechanical stirring. **9**⁵ (60 g, 0.22 mol) was added, the mixture turning brown. Refluxing was continued for 2 h. The solution was poured into dilute aqueous NaOH. Workup led to 23 g (40%) of tan prisms: mp 197–199 °C (lit. mp 197–199 °C).⁵

4-Bromo-6,7-methylenedioxy-2-oximino-1-hydrindone (11). To a warm solution of 10 g (39 mmol) of **10** in benzene was added 15 g (0.128 mol) of isoamyl nitrite and 10 mL of concentrated HCl. The mixture was stirred for 2 h at 50 °C. The bright yellow crystals which separated on cooling were collected, washed with methanol, and dried to give 9.48 g (85%) of bright yellow powder: mp 240 °C dec (lit. mp 240 °C dec).⁵

6-Bromo-3,4-methylenedioxyhomophthalic Acid (12). A solution of 14.2 g (0.05 mol) of 11 in 300 mL of cold aqueous NaOH was treated with 55 g (0.26 mol) of *p*-toluenesulfonyl chloride, and the mixture was stirred overnight in a cold water bath. To the resulting black solution, 10 g (0.25 mol) of NaOH was added, and the mixture refluxed 24 h. The cooled reaction mixture was acidifed and extracted with ethyl acetate. The organic extracts were washed with dilute acid, dried, filtered, and evaporated. The dark residue was dissolved in hot methanol, treated with charcoal, and filtered through a Celite pad. The methanol solution was concentrated to 75 mL and 300 mL of hot water was added. Most of the methanol was boiled off. The diacid crystallized upon cooling: 7.73 g (51%); mp 215 °C dec (lit. mp 215 °C dec).⁵

3,4-Methylenedioxyhomophthalic Acid (13). A solution of 6.0 g (20 mmol) of 12 in 200 mL of 1% aqueous NaOH was added to 200 g (0.26 mol of Na metal) of 3% Na/Hg. The temperature was maintained at 90 °C for 16 h. The mixture was filtered, concentrated, and acidified, and the product was collected. The aqueous solution was saturated with NH₄Cl and further extracted with ether. Recrystallization of the combined diacid fractions from hot water gave rise to 3.7 g (83%): mp 201–203 °C (lit. 203–204 °C).⁵

Dimethyl 3,4-Methylenedioxyhomophthalate (14). A solution

of 1.0 g (4.4 mmol) of 13 in 50 mL of methanol was saturated with HCl gas and refluxed 16 h. Workup and recrystallization from benzene produced 0.82 g (73%): mp 83–84.5 °C; $\nu_{\rm max}$ (CHCl₃) 1720 and 1735 cm⁻¹; high-resolution MS calcd for M⁺ C₁₂H₁₂O₆, *m/e* 252.0623, observed *m/e* 252.0610.

Piperonylidenemethylamine (15). A mixture of 100 g (0.68 mol) of piperonal and 150 g (1.93 mol) of 40% aqueous methylamine was stirred for 3 h and then extracted with ether. The ether solution was dried and evaporated. The residue was placed in a refrigerator where the product crystallized: white needles; 89 g (80%); mp 45–46 °C (lit. mp 46 °C).¹⁵

trans-1-Oxo-2-methyl-3-(3',4'-methylenedioxy)phenyl-4-(*N*-methyl)carboxamide-7,8-methylenedioxy-3,4-dihydroisoquinoline (16). A sodium methoxide solution was prepared by dissolving 1.5 g (65 mmol) of Na metal in 150 mL of methanol. Addition of 2.5 g (10 mmol) of 14 and 5.2 g (32 mmol) of 15 to this solution gave a pale yellow mixture which was heated to reflux. The mixture was saturated three times a day with methylamine gas, and a mercury seal was used to keep water and air out. Reflux was continued for 7 days, during which time the mixture turned opaque orange, and a yellow solid precipitated.

The solid was filtered off, washed with methanol, and dried. Recrystallization from methanol supplied 2.3 g (61%) of needles: mp 309–311 °C dec; ¹H NMR (TFA) δ 2.93 (3 H, s, NCH₃), 3.37 (3 H, s, NCH₃), 4.17 (1 H, d, J_{3.4} = 1 Hz, H-4), 5.42 (1 H, d, J_{3.4} = 1 Hz, H-3), 5.93 (2 H, s, OCH₂O), 6.22 (2 H, s, OCH₂O), 6.82 (1 H, d, J_{5.6} = 7 Hz, H-6), 7.07 (1 H, d, J_{5.6} = 7 Hz, H-5), and 6.64–6.77 (3 H, m, H-2', 5', 6'); ν_{max} (KBr) 1635 and 1655 cm⁻¹; λ_{max} (EtOH) 214 sh, 236 sh, 288, and 321 nm (log ϵ 4.44, 4.17, 3.80, and 3.65).

Anal. Calcd for $C_{20}H_{18}N_2O_6$: C, 62.82; H, 4.74. Found: C, 63.05; H, 4.94.

trans-1-Oxo-2-methyl-3-(3',4'-methylenedioxy)phenyl-4carboxy-7,8-methylenedioxy-3,4-dihydroisoquinoline (17). A suspension of 1.4 g (3.66 mmol) of amide 16 in 200 mL of aqueous 10% KOH was refluxed for 48 h under N₂. Workup and recrystallization from methanol supplied 0.95 g (70%): mp 249–254 °C dec; ¹H NMR (TFA) δ 3.47 (3 H, s, NCH₃), 4.25 (1 H, s, H-4), 5.40 (1 H, s, H-3), 5.98 (2 H, s, OCH₂O), 6.27–6.30 (2 H, d, OCH₂O), 6.66–6.95 (3 H, m, H-2', 5', 6'), 6.97 (1 H, d, J_{5.6} = 8 Hz, H-6), and 7.17 (1 H, d, J_{5.6} = 8 Hz, H-5).

Anal. Calcd for $C_{19}H_{15}NO_7$: C, 61.79; H, 4.09. Found: C, 61.43; H, 4.33.

Methyl Ester of 17. A solution of 0.4 g (1.1 mmol) of 17 in 150 mL of methanolic HCl was refluxed for 12 h. Workup and recrystallization from methanol generated 0.35 g (85%) of ester 18 as prisms: mp 213–214°C; ¹H NMR δ 3.12 (3 H, s, NCH₃), 3.72 (3 H, s, COOCH₃), 3.82 (1 H, d, J_{3,4} = 1.5 Hz, H-4), 5.08 (1 H, d, J_{3,4} = 1.5 Hz, H-3), 5.92 (2 H, s, OCH₂O), 6.17 (2 H, s, OCH₂O), 6.60 (1 H, d, J_{5,6} = 7.5 Hz, H-6), 6.83 (1 H, d, J_{5,6} = 7.5 Hz, H-5), and 6.55–6.80 (3 H, m, H-2', 5', 6'); ν_{max} (KBr) 1640 and 1730 cm⁻¹; high-resolution MS calcd for M⁺ C₂₀H₁₇NO₇, *m/e* 383.1004; observed *m/e* 383.1040.

Methyl Ester of trans-1-Oxo-2-methyl-3-(3',4'-methylenedioxy)phenyl-4-carboxymethyl-7,8-methylenedioxy-3,4-dihydroisoquinoline (19). A solution of 1.18 g (3.20 mmol) of acid 17 in 50 mL of chloroform was treated with 3 mL (35 mmol) of oxalyl chloride and stirred for 18 h in a flask equipped with a CaCl_2 drying tube. The solvent was evaporated to dryness, dry benzene was added, and the solvent was again evaporated. The residue was dissolved in chloroform and cooled in an ice bath. This solution was then added slowly to an ethereal diazomethane solution, and the mixture was left standing overnight in an ice bath. The precipitated crystals collected by filtration amounted to 1.05 g (83%) of crude diazoketone. A suspension of this material and 0.5 g of Ag₂O was heated to reflux in 200 mL of methanol for 1 h. The brown mixture was filtered through a Celite pad. The filtrate was evaporated to yield a brown residue. Crystallization from methanol gave 0.76 g (60%) of crystalline ester: mp 199-201 °C; ¹H NMR δ 3.07 (3 H, s, NCH₃), 2.67 (2 H, m, CH₂COOCH₃), 3.68 (3 H, s, OCH₃), 3.35 (1 H, m, H-4), 4.52 (1 H, d, $J_{3,4} = 1$ Hz, H-3), 5.82 (2 H, s, OCH₂O), 6.05 (2 H, s, OCH₂O), and 6.32–6.77 (5 H, m, ArH); ν_{max} (KBr) 1643 and 1720 cm⁻¹; λ_{max} (EtOH) 215 sh, 235 sh, 286, and 320 nm (log e 4.44, 4.20, 3.77, and 3.64); high-resolution MS calcd for M⁺ C₂₁H₁₉NO₇, m/e 397.1160; observed m/e 397.1159.

trans-1-Oxo-2-methyl-3-(3',4'-methylenedioxy)phenyl-4-carboxymethyl-7,8-methylenedioxy-3,4-dihydroisoquinoline (20). A suspension of 750 mg (1.89 mmol) of 19 in 100 mL of 10% aqueous KOH was refluxed for 3 h and the hot brown solution was treated with decolorizing carbon, filtered through a Celite pad, acidified with concentrated HCl, and extracted with chloroform. The extracts were washed with water and dried and the solvent was evaporated. The

Anal. Calcd for $C_{20}H_{17}NO_7$: C, 62.65; H, 4.47. Found: C, 62.72; H, 4.40.

trans-5,8-Dioxohexahydrosanguinarine (21). A solution of 5 g (35 mmol) of P_2O_5 in 50 g of methanesulfonic acid was warmed to 45 °C. To this solution was added 500 mg (1.31 mmol) of the above acid 20, and the mixture was stirred for 2 h while the temperature was maintained at 45 °C. The mixture was poured into ice water and extracted with chloroform. The organic solution was extracted with dilute aqueous NaOH and with water and dried, and the solvent was evaporated. The residue crystallized from ethanol: 210 mg (44%) as tan prisms; mp 277–280 °C dec; ¹H NMR (TFA) δ 3.38 (3 H, s, NCH₃), 2.55-4.08 (3 H, m, H-6 and H-13), 5.28 (1 H, d, $J_{13,14} = 11.5$ Hz, H-14), 5.33 (2 H, s, OCH₂O), 5.38 (2 H, s, OCH₂O), 6.85 (1 H, d, J_{11,12} = 8 Hz, H-12), 7.02 (1 H, s, H-1), 7.15 (1 H, d, $J_{11,12} = 8$ Hz, H-11), 7.53 (1 H, s, H-4); ν_{max} (CHCl₃) 1640 and 1675 cm⁻¹; λ_{max} (EtOH) 213, 237, 273, and 317 nm (log e 4.48, 4.60, 4.02, and 4.07).

Anal. Calcd for $C_{20}H_{15}NO_6$: C, 65.75; H, 4.14. Found: C, 65.71; H, 4.01

5-Hydroxy-8-oxohexahydrosanguinarine (22). A suspension of 100 mg (0.27 mmol) of the above keto lactam 21 and 100 mg (13 mmol) of NaBH₄ in 100 mL of isopropyl alcohol was stirred at room temperature for 16 h. The solvent was evaporated and water added to the residue. The mixture was acidified with councentrated HCl and extracted with chloroform. The organic extracts were washed with water and dried and the solvent was evaporated. The residue crystallized from methanol: 75 mg (74%) of white prisms; mp 281-283 °C dec; ν_{max} (KBr) 1620 and 3150–3600 cm⁻¹; λ_{max} (EtOH) 219 sh, 236 sh, 290, and 318 nm (log ϵ 4.40, 4.17, 3.80, and 3.59).

Anal. Calcd for C₂₀H₁₇NO₆: C, 65.39; H, 4.66. Found: C, 65.20; H, 4.76

Oxysanguinarine (23). A solution of 50 mg (0.14 mmol) of lactam alcohol 22 and 10 mg of p-toluenesulfonic acid in 50 mL of benzene was refluxed for 16 h. The solvent was evaporated and the residue was dissolved in chloroform. The solution was extracted with 5% aqueous NaHCO3 and dried, and the solvent was evaporated. The residue was subjected to preparative TLC using a 3:97 methanol-chloroform solvent system. A compound with an R_f 0.62, which was significantly higher than the R_f (0.29) of the starting lactam alcohol, was obtained. Recrystallization from ether gave 15 mg (30%), mp 347-349 °C dec, spectrally and chromatographically identical with oxysanguinarine: ν_{max} (CHCl₃) 1645 cm⁻¹; λ_{max} (EtOH) 241, 281 sh, 289, 331, 348, 370, and 385 nm (log ϵ 4.27, 4.61, 4.70, 4.17, 4.18, 4.06, and 4.02).

Acknowledgments. This project was supported by NIH research grant CA-11450, awarded by the National Cancer Institute, PHS/DHEW. The assistance of a departmental grant from the NSF toward the purchase of an NMR spectrometer is also acknowledged.

Registry No.—9, 56920-74-2; 10, 38699-84-2; 11, 66271-19-0; 12, 66271-20-3; 13, 66303-84-2; 14, 66271-21-4; 15, 63254-33-1; 16, 66271-22-5; 17, 66271-23-6; 18, 66303-85-3; 19, 66271-24-7; 20, 66271-25-8; 21, 66271-26-9; 22, 66271-27-0; 23, 548-30-1; piperonal, 120-57-0; methylamine, 74-89-5.

References and Notes

- For reviews on this subject, see G. A. Cordell and N. R. Farnsworth, *Heterocycles*, **4**, 393 (1976); *Lloydia*, **40**, 1 (1977).
 M. Onda, K. Abe, and K. Yonezawa, *Chem. Pharm. Bull.*, **16**, 2005 (1968).
- For the revised structure of bocconine, see: H. Ishii, K. Harada, T. Ishida, E. Ueda, and K. Nakajima, *Tetrahedron Lett.*, 319 (1975).
- E. Oeda, and K. Nakajima, *Tetrahedron Lett.*, 519 (1975).
 (3) For reviews on the chemistry and synthesis of benzophenanthridines, see: M. Shamma, "The Isoquinoline Alkaloids", Academic Press, New York, N.Y., 1972, p 315; M. Shamma and J. L. Moniot, "Isoquinoline Alkaloids Research, 1972–1977", Plenum Press, New York, N.Y., 1978, in press.
 (4) M. Shamma, R. W. Lagally, P. Miller, and E. F. Walker, Jr., *Tetrahedron*, 2655 (1965).
- 21, 3255 (1965). (5) R. D. Haworth, W. H. Perkin, Jr., and T. S. Stevens, *J. Chem. Soc.*, 1764 (1926).
- L. F. Fieser and M. Fieser, "Reagents for Organic Synthesis", Wiley, New York, N.Y., 1967, pp 892–894.
 D. M. Bailey, C. G. DeGrazia, H. E. Lape, R. Frering, D. Fort, and T. Skulan, (6)
- (7) *J. Med. Chem.*, **16**, 151 (1973). Reference 6, pp 880-881.
- (8)(9) W. H. Hartung and F. Crossley, "Organic Syntheses". Collect. Vol. 2, Wiley, New York, N.Y., 1943, p 363.
 (10) A. F. Ferris, G. E. Johnson, and F. E. Gould, J. Org. Chem., 25, 1813
- 1960).
- (11) D. J. Pasto and C. R. Johnson, "Organic Structure Determination", Prentice-Hall, Englewood Cliffs, N.J., 1969, pp 183-186.
- (12) P. E. Eaton, G. R. Carlson, and J. T. Lee, J. Org. Chem., 38, 4071 (1973).
- (13) D. B. MacLean, D. E. F. Gracey, J. K. Saunders, R. Rodrigo, and R. H. F. Manske, *Can. J. Chem.*, **47**, 1951 (1969). For an alternate method for achieving this transformation see: A. S. Bailey and R. Robinson, *J. Chem.* Soc., 1375 (1950); A. S. Bailey, R. Robinson, and R. S. Staunton, ibid., 2277 (1950).
- (14) E. Spath, F. Schlemmer, G. Schenk, and A. Gempp, *Ber. Dtsch Chem. Ges.*, *B*, **70**, 1677 (1937).
- (15) J. R. A. Pollock and R. Stevens, Ed., Dictionary of Organic Compounds", Oxford University Press, New York, N.Y., 1965, p 2188.

Chemistry of Chelocardin. 3.1 Structure and Synthesis of Isochelocardin

Edith Bernstein, Daniel T. W. Chu,* Stuart N. Huckin, and David L. Garmaise

Abbott Laboratories, Limited, Montreal, Quebec, Canada H3C 3K6

Richard S. Egan, Thomas J. Perun, William Rosenbrook, Jr., and Ronald E. Carney

Abbott Laboratories, North Chicago, Illinois 60064

Received January 17, 1978

Isochelocardin (2), a minor component of the chelocardin fermentation, was shown to be a condensation product of two molecules of chelocardin. Carbobenzoxyisochelocardin acethydrazone (9) was synthesized by treatment of carbobenzoxychelocardin with chelocardin acethydrazone, thus confirming the assigned structure. The synthesis of isochelocardin itself is also described.

During the isolation of chelocardin (1),^{2,3} a potent broadspectrum antibiotic produced by Nocardia sulphurea (NRRL-2822), a contaminant which we designated as isochelocardin, was noted to be present and was subsequently isolated as a hydrochloride salt after chromatographic separation. This compound was present in the isolated chelocardin in proportions ranging from 1 to 3%. In view of the potential

